Preclinical Modeling of Laser Interstitial Thermal Therapy in Murine Brain Tumors for Translational Immuno-oncology



Duke Neurosurgery

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INTRODUCTION

- > Cytoreduction is a cornerstone of the management of intracranial tumors.
- > Following laser interstitial thermal therapy (LITT), ablated tumor tissue remains in situ, allowing for an anti-tumor immune response.
- The post-LITT immune response has not been well characterized previously.

Objective: To develop a murine model of stereotactic laser ablation for identification of combinatorial treatment approaches.

METHODS

Our murine LITT system is depicted in **Figure 1**. Following creation of a craniotomy, a 1064 nm diode-based Nd:YAG laser (Neurolase, Monteris Medical) was used to thermally ablate normal brain tissue or CT-2A tumors in C57BL/6 mice. Ablations were performed at + 2 ML, + 1 AP, -3 DV using a stereotactically inserted 400 µm core fiber (ThorLabs); simultaneous temperature recording was performed 2 mm posterior to the ablation site using a needle thermocouple (MT-29, Physitemp). T2 sequences with TE of 54 ms and TR 3000 ms were acquired using a 7T Bruker BioSpec 70/20 small animal MRI using Bruker Paravision 6 software (Center for In Vivo Microscopy, Duke University School of Medicine). Formalin-fixed, paraffin-embedded mouse brains were stained and imaged using a Keyence BZ-X800.

RESULTS

To identify a well-tolerated thermal dose, we ablated brains from nontumor bearing mice at 1, 1.5, or 2 W for 30 – 90 seconds each, finding that 1 W for 60 s did not cause post-LITT mortality (Fig. 2A, red points = 24h mortality). Notably, temperature measurements at lower power settings appeared more consistent overall (Fig. 2B - 2D). On H&E staining at 24h, ablations at lower powers and times produced a controlled ablation that did not char tissue (Fig. 2E). These data suggest higher, mortality-associated doses (1.5 W for 90s and beyond) produced large cavitary lesions associated with tissue vaporization and charring, causing inaccuracy in temperature measurement (Fig. 2F).

Although real-time MR thermometry is not feasible for high-throughput animal studies, we used serial T2-weighted MRI imaging to accurately track CT-2A tumor growth (Fig. 3A,n = 5 mice) and identify a safe lesion size for ablation, selecting 10 days post-implantation for subsequent experiments (Fig. 3B). Based on tumor diameter at these timepoints, we selected 2 mm posterior to the implantation site to monitor temperature at the lesion edge to mitigate damage to surrounding tissue. (Fig. 3C).









without Real-Time MR Thermometry time point. All values are means and standard deviations.





image, inset = 10x magnification, scale bar = 250μ m.

Figure 1. Murine LITT System needle thermocouple and fiber optic were threaded into a 1 mL syringe held in a stereotactic probe holder (left panel) and inserted into a burr-hole craniotomy (middle and right panels) to a depth of 3 mm at +2 ML. +1 AP.

RESULTS

A. Serial T2-weighted MRI of CT2A brain tumors at day 7, 10, and 13 post-implantation. N = 5 mice, scale bar = 5 mm. B-C. Segmented tumor volumes and perpendicular diameters at each



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imaging timepoint. * = P < 0.05, Student's t-test.

Figure 2. Identification of a Non-Morbid Thermal Dose for

A. Quantification of the peak B-D. and time:temperature relationship in °C for varying thermal dosages to naive mouse brains based on thermocouple recordings. N = 3animals per group, all graphs plotted as means with standard deviations. **E.** H&E images at 10x magnification of ablated brains at varying thermal doses 24 hours following surgery, scale bar = 250 µm F. Stitched whole brain images of mice treated at each power level showing gradual progression from coagulative necrosis to cavitary, charred tissue post-LITT.

We next implanted CT-2A tumors for ablation with the selected dose, following mice for survival between the LITT and sham-ablated groups. A subset of mice underwent imaging at 1, 3, and 7 days post-LITT. The peak temperature at the lesion edge achieved with intratumoral ablations was 43.79 ± 2.99 °C (Fig. 4A-B), slightly higher than that of naïve brain tissue at the same thermal dose (41.43 \pm 0.71 °C). Median survival for sham-operated mice was 25.5 days vs 28 days for LITTtreated mice, P = 0.0172, (Fig. 4C). T2-weighted MRI scans revealed hyperintense lesions consistent with coagulated, hemorrhagic portions of treated tumor at post-LITT days 1 and 3 (Fig. 4D). Interestingly, tumor did not appear to regrow in these spaces at post-LITT day 7, with flow voids remaining in their place. H&E staining of mice at sacrificed at clinical endpoints revealed persistent hemorrhagic and fibrotic lesions along the fiber tract (Fig. **4E**). These findings were also seen on studies employing bioluminescence imaging (Fig. 5A- 5C).

CONCLUSIONS

- > Our model successfully recapitulates the essential clinical features of LITT: a cytoreductive survival benefit, lesion expansion on MRI, and focal necrosis with hemorrhagic vasculature on H&E.
- > Our model can be used in a modular fashion to effects laser-induced investigate the of hyperthermia on syngeneic glioma or brain metastasis cell lines, as well as in patient-derived xenografts or murine epilepsy paradigms.

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dural puncture) versus LITT-treated mice. Median overall survival = 25.5 vs 28 days, log-rank test, P = 0.0172 **D.** Serial T2-weighted MRI of mice treated with LITT at post-LITT day 1, 3, and 7. Scale bar = 5 mm. **E**. Representative H&E staining of a LITT-treated mouse at clinical endpoint. Left panel = stitched whole brain